

# Fish Health Inspection Report

**Company:** Blackwater Creek Koi Farms  
**Facility :** Blountstown Facility  
**Location:** 24946 State Road 69 NE  
 Blountstown, FL 32424  
 USA

**Site Manager:** Joe Pawlak  
**Phone:** (352) 357-4563  
**Water Source:** Well  
**Water Treatment:** None

**Current Inspection:** 08-Feb-12  
**Prior Inspections:**

**Type of Fish Examined:** Hatchery

**Lab Accession:** M12020902

Species	Lot ID	Age	Number in Lot	Eggs (E) or Fish (F) Origin	Sample Date(s)	*Pathogens - Methods and Results								
						Viruses				Bacteria		Parasites		
						IPNV	KHV	SVCV	VHSV	VNNV	BF	BRM	HSP	B. ach.
<i>Carassius auratus</i> goldfish	M12-089	~12 mo	10,000	(E) Blackwater Creek Koi Farms (FL)	08-Feb-12	P24B 15 neg		P42B 15 neg	P42B 15 neg					
<i>Cyprinus carpio koi</i> koi	M12-090	~12 mo	200,000	(E) Blackwater Creek Koi Farms (FL)	08-Feb-12	P24B 135 neg	P24B 135 neg	P42B 135 neg	P42B 135 neg					

**Notes:** \* See other side of sheet for explanations of Pathogens - Methods and Results coding  
 All lots were tested according to World Organization for Animal Health (OIE) "Manual of Diagnostic Tests for Aquatic Animals" (2009) protocols. At the time of collection, all fish were visually examined by Kathleen Hartman, DVM (USDA/APHIS/VS) for the presence of gross clinical signs consistent with Epizootic Ulcerative Syndrome (EUS). None of the fish examined from lot M12-089 or M12-090 had clinical signs of EUS.

**Samples Collected By:** Sherri Kasper, DVM

**Affiliation:** Northwood Animal Hospital

**Telephone:** (850) 385-8181

**Client Reference #:**

**Inspecting Biologist:**

*William Keleher*

William R. Keleher, Jr., Fish Health Official



\* Notes are located on the last Page of this report

GEN R022

Report Issued: 3/21/2012

BWCBF.I.M12020902F.pdf

# FOOTNOTES:

## **PATHOGEN ABBREVIATIONS**

IPNV Infectious Pancreatic Necrosis virus  
IHNV Infectious Hematopoietic Necrosis virus  
VHSV Viral Hemorrhagic Septicemia virus  
VEN Viral Erythrocytic Necrosis virus  
HPV Herpesvirus salmonis  
SVCV Spring Viremia Carp virus  
YTV Yamame Tumor virus  
EEV Epizootic Epitheliotropic virus  
ISAV Infectious Salmon Anemia virus  
EHNV Epizootic Haematopoietic Necrosis virus  
BF Aeromonas salmonicida  
BRM Yersinia ruckeri  
BKD Renibacterium salmoninarum  
WD Myxobolus cerebralis  
CS Ceratomyxa shasta  
PKD Proliferative Kidney Disease  
HSP Heterosporis spp.  
B. ach. Bothriocephalis acheilognathi (Asian Tapeworm)

In lots of fish less than one year of age, the age is listed in arabic numerals followed by mo. for month; for fish older than one year, the age is expressed in arabic numerals followed by a plus sign to indicate "older than".

Findings are reported in columns from top to bottom for each lot as follows: number of fish examined; methods used; results. Positive results include the number of positive individuals (or pools).

**RESULTS ARE REPORTED AS (-) IF NEGATIVE AND AS # + / # SAMPLED IF POSITIVE.**

**FOR BKD, APPROXIMATE LEVELS OF INFECTION ARE ALSO REPORTED (e.g., 10/ 50 fish)**

## **VIRAL PATHOGENS:**

### **First letter = sampling method**

A = whole fry homogenates( minus head, tail, yolk sac if present)  
B = whole visceral homogenates  
C = kidney/spleen  
D = ovarian fluids  
E = kidney/spleen/heart  
F = kidney/spleen/liver  
G = kidney/spleen/heart/liver/pyloric caeca/gill  
H = kidney/spleen/swim bladder  
I = kidney/spleen/heart/liver  
J = brain/eye  
K = kidney/spleen/pyloric caeca/gill  
L = kidney/spleen/heart/swim bladder  
M = kidney/spleen/liver/swim bladder  
N = kidney/spleen/heart/liver/swim bladder  
O = kidney/spleen/heart/pyloric caeca/gill  
P = kidney/spleen/heart/liver/gill

### **Numbers = continuous cell lines used**

1 = RTG-2 (rainbow trout gonad)  
2 = CHSE-214 (chinook salmon embryo)  
3 = FHM (fathead minnow)  
4 = EPC (epithelioma papillosum cyprini)  
5 = BF-2 (bluegill fry)  
6 = SHK 1/3 (salmon head kidney)  
7 = ASK (atlantic salmon kidney)  
8 = SSN-1 (striped snake head )  
9 = KF-1 (koi fin)  
10 = CCO (Channel Catfish Ovary )

### **Last letter = Pooling of samples**

A = individual fish  
B = five fish pools  
C = Other \_\_\_\_\_

## **PROTOZOAN/PARASITIC PATHOGENS:**

A = Digestion method  
B = Plankton centrifuge method  
C = Examination of stained smear  
D = Gross Examination  
E = PCR (Polymerase Chain Reaction)  
F = Microscopic Examination

## **BACTERIAL PATHOGENS:**

Encoded as follows:

### **First letter = Health of fish sampled**

A = Live, healthy fish  
B = Moribund fish  
C = Mortalities

### **Number = Material sampled**

1 = kidney  
2 = hindgut  
3 = lesion  
4 = gill  
5 = ovarian fluid  
6 = seminal fluid  
7 = Other \_\_\_\_\_

### **Last letter = technique used for:**

#### **Primary Isolation**

A = Standard culture medium TSA/BHI  
B = Cytophaga agar  
C = KDM2/SKDM2  
D = Kidney smear/impression

#### **Presumptive Diagnosis**

E = Gram stain, kidney smears (BKD)  
F = Standard biochemical/physical testing  
G = Giemsa stain

#### **Confirmatory diagnosis**

H = Slide agglutination  
I = Direct fluorescent antibody test  
J = Indirect fluorescent antibody test  
K = ELISA  
L = Immunodot  
M = Fluorescent immunoassay  
N = PCR

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 US

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**Phone:** (352) 357-4563  
**Water Source:** Well  
**Water Treatment:** None

**Current Inspection:** 28-Nov-12      **Prior Inspections:** 08-Feb-12

**Type of Fish Examined:** Hatchery

**Lab Accession:** M12112901

Species	Lot ID	Age	Number in Lot	Eggs (E) or Fish (F) Origin	Sample Date(s)	*Pathogens - Methods and Results									
						Virus							Parasite		
						EHN	INH	IPNV	ISAV	KHV	RSIV	SVCV	VHSV	EUS	Gyro
<i>Cyprinus carpio koi</i>	M13-004	~6-12 mo	25,000	(E) Blackwater Creek Koi Farms (FL)	28-Nov-12			P24B		P94B		P42B	P42B	D	
								120		120		120	120	120	
								neg		neg		neg	neg	neg	
<i>Carassius auratus goldfish</i>	M13-005	~6-12 mo	8,000	(E) Blackwater Creek Koi Farms (FL)	28-Nov-12			P24C				P42C	P42C	D	
								30				30	30	30	
								neg				neg	neg	neg	

Notes: \* See other side of sheet for explanations of Pathogens - Methods and Results coding  
 All lots were tested according to World Organization for Animal Health (OIE) "Manual of Diagnostic Tests for Aquatic Animals" (2009) protocols. At the time of collection, all fish were visually examined by Sherri Kasper, DVM for the presence of gross clinical signs consistent with Epizootic Ulcerative Syndrome (EUS).

**Samples Collected By:** Sherri Kasper , DVM  
**Affiliation:** Northwood Animal Hospital  
**Telephone:** (850) 385-8181  
**Client Reference #:**  
**Inspecting Biologist:** *William Keleher*  
 William R. Keleher, Jr., Fish Health Inspector

# FOOTNOTES:

## PATHOGEN ABBREVIATIONS

EHN	Epizootic Hematopoietic Necrosis virus	
IHN	Infectious Hematopoietic Necrosis virus	Infectious
IPNV	Pancreatic Necrosis virus	
ISAV	Infectious Salmon Anemia virus	
KHV	Koi Herpes virus	
RSIV	Red Sea Bream iridovirus	
SVCV	Spring Viremia of Carp virus	
VHSV	Viral Hemorrhagic Septicemia virus	
EUS	Epizootic ulcerative syndrome	
GYRO	Gyrodactylus salaris	

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N = kidney/spleen/heart/liver/swim bladder  
O = kidney/spleen/heart/pyloric caeca/gill  
P = kidney/spleen/heart/liver/gill

### Numbers = continuous cell lines used

1 = GF-1 (grunt fin)  
2 = CHSE-214 (chinook salmon embryo)  
3 = FHM (fathead minnow)  
4 = EPC (epithelioma papillosum cyprini)  
5 = BF-2 (bluegill fry)  
6 = CCO (channel catfish ovary )  
7 = ASK (atlantic salmon kidney)  
8 = SSN-1 (striped snake head )  
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### Last letter = Pooling of samples

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## PARASITIC PATHOGENS

### Encoded as follows:

A = digestion method  
B = plankton centrifuge method  
C = examination of stained smear  
D = gross examination  
E = PCR  
F = microscopic examination